



Toxicity of carbon dioxide and its relationship to tobacco smoke

Journal:	<i>Critical Reviews in Toxicology</i>
Manuscript ID:	Draft
Manuscript Type:	Review
Date Submitted by the Author:	n/a
Complete List of Authors:	Guais, Adeline; Biorebus Brand, Gérard; Université de Franche-Comté, Laboratoire de Neurosciences; INRA, Centre des Sciences du Goût et de l'Alimentation (CSGA) Jacquot, Laurence; Université de Franche-Comté, Laboratoire de Neurosciences Karrer, Mélanie; Université de Franche-Comté, Laboratoire de Neurosciences Grévillet, Georges; CNRS - ENSIC, Laboratoire des Sciences du Génie Chimique Molina, Thierry; Université Paris Descartes, AP-HP Hôtel-Dieu Bonte, Jacques; Biorebus Regnier, Mireille; Ecole Polytechnique, Laboratoire d'informatique Schwartz, Laurent; Ecole Polytechnique, Laboratoire d'informatique;

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Keywords:	Acute toxicity, chronic toxicity, blood pH, metabolism, cancer, lung, heart, teratogenicity, cigarette, inflammation, central nervous system, hypercapnia, reproduction, PaCO ₂ , carcinogenesis
-----------	---

SCHOLARONE™
Manuscripts

For Peer Review Only

Toxicity of carbon dioxide and its relationship to tobacco smoke

Adeline Guais¹, Gerard Brand^{2,3}, Laurence Jacquot³, Mélanie Karrer³, Georges Grévillet⁴,
Thierry Jo. Molina⁵, Jacques Bonte¹, Mireille Regnier⁶, Laurent Schwartz^{6,7*#}.

¹ Biorébus, Paris, France.

² Centre des Sciences du Goût et de l'Alimentation (CSGA) - Dijon

³ Laboratoire de Neurosciences - Université de Franche-Comté, Besançon, France.

⁴ Laboratoire des Sciences du Génie Chimique CNRS – ENSIC, Nancy, France.

⁵ Université Paris Descartes, AP-HP Hôtel-Dieu, Paris, France.

⁶ Ecole Polytechnique, Laboratoire d'informatique, Palaiseau, France.

⁷ AP-HP Hôpital Pitié-Salpêtrière, Service de radiothérapie, Paris, France.

* to whom request for reprints should be sent laurent.Schwartz@polytechnique.fr

Corresponding author:

Dr Laurent Schwartz, Service de Radiothérapie Hôpital Pitié-Salpêtrière, bd. de l'Hôpital,
75013 Paris, France.

e-mail: laurent.schwartz@polytechnique.edu

tel: +33 681899030 fax: +33 140700130

Key words:

Acute toxicity, chronic toxicity, blood pH, metabolism, cancer, lung, heart, teratogenicity,
cigarette, inflammation, central nervous system, hypercapnia, reproduction, PaCO₂,
carcinogenesis.

Abstract

The toxicity of carbon dioxide has been established for close to a century. A number of animal experiments have explored both acute and long-term toxicity with respect to the lungs, the cardiovascular system and the bladder, showing inflammatory and possible carcinogenic effects. Carbon dioxide also induces malformations and probably reduces fertility. As smokers are exposed to a high level of carbon dioxide (13.5%) that is about 500 times the level in normal air, the aim of this paper is to review the physiological and metabolic mechanisms resulting from CO₂ inhalation and supporting the hypothesis that carbon dioxide plays a major role in the long term toxicity of tobacco smoke.

Table of contents:

1.	Perturbation of acid/base balance by carbon dioxide.....	5
a-	Acute respiratory acidosis	5
b-	Chronic respiratory acidosis.....	6
2.	Metabolic effects of carbon dioxide.....	8
a-	CO ₂ implication in cellular metabolism	8
b-	Alteration of in vivo metabolism caused by CO ₂	9
3.	Pulmonary toxicity of carbon dioxide.....	11
a-	Respiratory function	11
b-	Acute and chronic lung toxicity	12
4.	Effects of carbon dioxide on cardiovascular function.....	14
5.	Effects of carbon dioxide on central nervous system and neuroendocrine function.....	15
6.	Alteration of the reproductive capacity by carbon dioxide.....	16
7.	Teratogenicity of carbon dioxide	16
8.	Carcinogenic potential of carbon dioxide	17
a-	In vitro alteration of cell fate by CO ₂ exposure.....	17
b-	In vivo carcinogenicity of carbon dioxide.....	19
9.	Does carbon dioxide contribute to cigarette smoke toxicity?	21

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Carbon dioxide (CO₂) is naturally present in the atmosphere where its concentration varies from 0.03 to 0.06 % (vol/vol, equivalent to 0.2 mmHg to 0.4 mmHg) (Keeling, 1995). Its regularly increasing concentration contributes to the greenhouse effect (Bertoni, 2004) and the acceleration of global warming (Cox, 2000). The average indoor concentration of CO₂ is 0.08% to 0.1% (National Research Council [NRC], 2008). The maximal acceptable concentration has been defined between 0.5 and 3%, depending on duration of exposure. At normal temperature and pressure, carbon dioxide is an odorless, colorless, and heavier than air gas, with a faintly pungent odor (Shusterman, 1997). CO₂ is widely used in industries, especially in agro-productions for conserving, cooling and medical applications. It is also known to be produced during combustion, putrefaction and fermentation.

In air, carbon dioxide is a very stable and non-flammable compound. As CO₂ is soluble in water, it can react to form carbonic acid (H₂CO₃). Dissolved carbon dioxide in the water undergoes hydration according to the following reaction: CO₂ + H₂O ↔ H₂CO₃ ↔ H⁺ + HCO₃⁻. This reaction can interfere with the acid-base balance: pH = pK + log [HCO₃⁻/CO₂] (Henderson-Hasselbach equation).

Carbon dioxide is a normal constituent of the human body arising from cellular respiration (National Institute for Occupational Safety and Health [NIOSH], 1976). Carbon dioxide diffuses from cells into the surrounding capillaries and is carried by the blood either bound to hemoglobin or dissolved as carbon dioxide, carbonic acid, or bicarbonate ion (Baggot, 1982). A minor amount of CO₂ can be bound to plasma proteins to form carbamino compounds.

Carbon dioxide is synthesized in the body and its partial pressure under normal conditions in pulmonary capillary blood (almost 7% or 46 mm Hg) is greater than that in alveolar air (6% or 40 mm Hg). The gas is exchanged freely through the alveolar membrane

1
2
3 and is thus released from the lungs by diffusion because of the concentration gradient
4 existing between the blood and the air in the alveoli. Its free diffusion through the lipid cell
5 membranes allows it to be one of the main regulators of intracellular pH acting as a stimulant
6 or a brake in numerous cellular processes. Due to its free diffusion through tissue
7 membranes, the toxicological effects of carbon dioxide appear very rapidly and are mainly
8 observed on the blood pH, lungs, heart and central nervous system.
9
10
11
12
13
14
15
16
17
18
19

20 **1. Perturbation of acid/base balance by carbon dioxide**

21
22
23 An increase of the partial pressure of CO₂ (pCO₂) delivered to the lungs, i.e.,
24 hypercapnia, induces an increase of pCO₂ in the alveoli. Because carbon dioxide freely
25 diffuses through the alveolar membrane to the blood, it results in an increase of the CO₂
26 tension in arterial blood (PaCO₂). This increase in PaCO₂ results in turn in an acute or
27 chronic respiratory acidosis.
28
29
30
31
32
33
34

35 **a- Acute respiratory acidosis**

36
37
38 In acute respiratory acidosis, the PaCO₂ is elevated above the upper limit of the
39 reference range (i.e., >6.75% or 45 mm Hg) resulting in acidosis (i.e., pH <7.35). This acute
40 hypercapnia can be compensated for in two steps. The initial response is cellular buffering
41 that occurs within minutes to hours (NIOSH, 1976). Cellular buffering elevates plasma
42 bicarbonate (HCO₃⁻) only slightly, approximately 1 mEq/L for each 0.15% increase (10 mm
43 Hg) in PaCO₂. The second step is renal compensation that occurs over 3-5 days: renal
44 excretion of carbonic acid and bicarbonate reabsorption are increased. In renal
45 compensation, plasma bicarbonate increases 3.5 mEq/L for each increase of 0.15% (10 mm
46 Hg) in PaCO₂.
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4 For acute respiratory acidosis, the expected change in pH with respiratory acidosis
5
6 can be estimated by the following equation: $\text{change in pH} = 0.008 \times (40 - \text{PaCO}_2)$ (Smith,
7
8 2005). This means that, from a (normal) pH of 7.4 for 6 % PaCO₂ and 0.03% CO₂ in the
9
10 atmosphere, the pH could fall to 6,65 when PaCO₂ is increased to 20%. In guinea pigs
11
12 exposed to 15% CO₂ during 1 hour (acute response), the PaCO₂ value was reported to be
13
14 17.8 % (119 mmHg) (Schaefer, 1964a).
15
16
17
18
19
20

21 **b- Chronic respiratory acidosis**

22
23 In chronic respiratory acidosis, the value of the pH is subnormal secondary to renal
24
25 compensation and an elevated concentration of serum bicarbonate. In rats exposed to 10% or
26
27 15% CO₂ during 11 days (chronic acidosis), Carter et al. measured a plasma CO₂ content of
28
29 45 and 52 mEq/l respectively and estimated the PaCO₂ values at 15% (102 mmHg CO₂) and
30
31 22% (148 mmHg CO₂) respectively (Carter, 1959).
32
33
34

35 In guinea pigs exposed to 15% CO₂ (about 300 time the normal air level) for 73 days
36
37 (Schaefer, 1961), the uncompensated acidosis period (first variations noticed within one day)
38
39 is characterized by a decline of extracellular and urine pH, inorganic phosphorus plasma
40
41 concentration, and an increase of the calcium plasma concentration and urine inorganic
42
43 phosphorus. During the compensated period, the extracellular pH returns to normal but
44
45 plasma calcium is still elevated and inorganic phosphorus low level is maintained even after
46
47 20 days of exposure. This effect on calcium and inorganic phosphorus was associated with
48
49 renal calcification after 48 hours of exposure. In rats exposed to 10 or 15% CO₂ for 11 days
50
51 (Carter, 1959), an increase in urine excretion of ammonia and acidic substances was
52
53 observed. During the first two days (acute response), the ammonia and titratable acid
54
55 excretion was almost twice the normal values, and the urine pH value was around 6.2 (10%
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

CO₂). Potassium and chloride ions were significantly increased during the first days of exposure.

Body adaptation to chronic high carbon dioxide level is dependent of the concentration administered: below 3% CO₂ the compensatory mechanisms occur more slowly. Volunteers were exposed to 1.5% CO₂ over a period of 42 days and acid-base balance and changes in electrolyte metabolism were studied (Schaefer, 1964b). During the first 23 days, a slight uncompensated respiratory acidosis was present followed by a compensated acidosis. Interestingly, arterial CO₂ tension increased by 5 mmHg (0.75%) during exposure and remained at this level during the first nine days of recovery in air. Several other studies were performed in men with low levels of increased carbon dioxide exposure (1.5 to 3%) in order to mimic living conditions in submarines or in space. Interestingly, although there were some minor modifications of the pH and serum level of the electrolytes, the experimental conditions were well tolerated (for review see Glatte, 1967a; Glatte, 1967b; Schaefer, 1979).

Guinea pigs were exposed to chronic intermittent high carbon dioxide level (8 hours per day during 7 days, 15% CO₂). While animals exposed to constant CO₂ (15% CO₂ during 7 days) displayed a 3-day-long uncompensated phase and then stabilized (pH: 7.37±0.035), animals exposed to intermittent CO₂ could not compensate for the respiratory acidosis, and the pH value was decreased by 0.26 (pH: 7.111±0.07) (Schaefer, 1968). Similarly, Schaefer et al (Schaefer, 1970) investigated the acid-base and electrolyte responses to intermittently increased carbon dioxide concentration (concentration increasing up to 3 % CO₂, 15 hours/day during 5 days) in human beings. The author reported doubling of urine volume on the fourth and fifth days. This increase in urine volume was accompanied by increases in organic acids, titratable acidity and ammonia, reflecting the elimination of the accumulated carbon dioxide by the kidneys.

1
2
3 Thus chronic carbon dioxide high tension exposure causes a raise of extracellular acidity
4 that is compensated within days (constant 10% or 15%) or weeks (constant 1.5 or 3%).
5
6 However intermittent exposure to carbon dioxide does not allow the compensation
7
8 mechanisms to be active.
9
10
11

12 13 14 15 **2. Metabolic effects of carbon dioxide**

16 17 18 **a- CO₂ implication in cellular metabolism**

19
20
21
22 The effects of carbon dioxide on metabolims have been poorly investigated. Warburg,
23 Posener and Negelein (Warburg, 1924) performed the first work on the metabolic effects of
24 carbon dioxide and they demonstrated the sensitivity of anaerobic glycolysis in a tumor to
25 the concentration of the carbon dioxide-bicarbonate buffer system. In 1943, Craig and
26 Beecher (Craig, 1943) demonstrated that the metabolism in the retina is sensitive to the
27 concentration of the carbon dioxide-bicarbonate buffer system. Increasing the carbon dioxide
28 from 1% to 5% at constant pH increases almost two-fold both glycolysis and cellular
29 respiration. Increasing the carbon dioxide at constant pH from 5% to 20% had no effect on
30 glycolysis, but depressed respiration. This was later confirmed in several studies on different
31 normal and cancer cells (for review see Goldsmith, 1970).
32
33
34
35
36
37
38
39
40
41
42
43
44
45

46 CO₂ is a product of oxidative metabolism but CO₂ and its by-product HCO₃⁻ is also a
47 substrate for important biochemical reactions occurring, for example, in the mitochondria
48 (Lahiri, 2003). CO₂ takes part into two types of reactions controlling respiration in animals:
49 the formation and transport of H⁺ (by reversible hydration of CO₂ and by formation of
50 carbamates from the NH₂ group of proteins) and the stimulation of metabolism.
51
52
53
54
55
56
57

58 HCO₃⁻ is required in at least three metabolic pathways in the mitochondria of the liver.
59 Mitochondria are impermeable to HCO₃⁻, so that the required anion must be provided by the
60

1
2
3 hydration of CO₂ which can diffuse easily across the membrane. Hydration of CO₂ is the
4
5 rate limiting factor for these three metabolic pathways. HCO₃⁻ is involved in the formation of
6
7 malonyl-CoA (enzyme: acetyl-CoA carboxylase) used for the production of fatty acids
8
9 components of cell membranes. CO₂ is needed for the conversion of pyruvate to
10
11 phosphoenolpyruvate during gluconeogenesis (enzyme: pyruvate carboxylase). Carbon dioxide
12
13 is also required for the synthesis of carbamoyl phosphate (enzyme: carbamyl phosphate
14
15 synthetase I). This is known to be the entry in the urea cycle and the regulated reaction of the
16
17 pyrimidine biosynthesis.
18
19
20
21

22 It was demonstrated *in vitro* that the inhibition of a specific liver mitochondrial
23
24 carbonic anhydrase isoenzyme, the catalyser allowing a rapid conversion of CO₂ into
25
26 bicarbonate, reduces the formation of glucose, urea and fatty acids in hepatocytes (Forster,
27
28 2000). Furthermore, raising the CO₂ concentration (up to approximately 8.5%) increases the
29
30 carboxylation of ¹³C labeled pyruvate independently of pH (Ono, 1996).
31
32
33
34
35

36 **b- Alteration of in vivo metabolism caused by CO₂**

37
38
39 Douglas and al. demonstrated that, in guinea pigs, exposure to 1% CO₂ during six
40
41 weeks did not alter weight evolution as compared to controls (Douglas, 1979). Schaefer et al
42
43 (Schaefer, 1971) studied the effect of long-term exposure of guinea pigs to higher tensions of
44
45 carbon dioxide with respect to several aspects of metabolism. With exposure to 1.5% CO₂,
46
47 they observed that the guinea pigs lost weight for about 25 days. The animals then start to
48
49 regain weight but at a slower rate than that of the controls (2.2 g/day versus 4.75 g/day).
50
51 During long term 3% CO₂ exposure, approximately 35 days are required for the weight of
52
53 the animals to start increasing above the initial level. During exposure to 15% CO₂, a 10%
54
55 loss of weight occurred during the first two days. At day 20 the rodents started to gain weight
56
57 for about 20 days to about 50 days.
58
59
60

1
2
3
4 *In vivo* CO₂ exposure also affects the expression or activity of certain metabolic
5 enzymes (Schaefer, 1971). Exposure of guinea pigs to 15% CO₂ for seven days results in a
6 striking but transient increase in plasma levels in GOT (glutamic oxaloacetic transaminase)
7 and GPT (glutamic pyruvic transaminase). After seven days of prolonged exposure, the
8 concentrations of these two enzymes return to the initial values. These variations follow the
9 pH changes corresponding to the uncompensated phase of respiratory acidosis. The activity
10 of other serum enzymes such as lactate dehydrogenase, malate dehydrogenase, isocitrate
11 dehydrogenase and cholinesterase increases significantly during the first three days of
12 exposure (uncompensated phase) and return to control level after seven days.
13
14

15
16
17
18
19
20
21
22
23
24
25 Histopathology analyses showed that prolonged exposure to CO₂ (3% during 7 days)
26 causes the depletion of glycogen vacuoles and an increase in fat vacuoles in guinea pigs
27 (Schaefer, 1971). After three weeks of exposure to 3% CO₂ and subsequent recovery for one
28 day breathing normal air, glycogen is again synthesized. These functional changes point to
29 important changes in fat metabolism caused by hypercapnia. Acidosis is known to inhibit
30 lipolysis (Poyart, 1968), and one could therefore expect an increase in fat since it would not
31 be easily mobilized. It is noteworthy that both guinea pigs and rats when exposed to even
32 low levels of CO₂ (3%) exhibit similar changes in glycogen and fat vacuolization. That
33 would seem to suggest that modifications of fat metabolism are of special significance in
34 hypercapnia. Lipid accumulation during chronic hypercapnia (15%) shows a specific pattern
35 for different organs. Fat content in muscle is increased only during the first two days, that of
36 lungs during the period from three to seven days, while the lipid content of the liver is
37 greatly elevated throughout the exposure period.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53

54
55 Several *in vitro* or *in vivo* studies have demonstrated that acidosis inhibits lipolytic
56 activity (Triner, 1965; Hollidge-Horvat, 1999). Adrenaline-induced lipolysis and
57 calorogenesis is inhibited in dogs when breathing a mixture of 10% CO₂ and 25% O₂ in N₂
58
59
60

1
2
3 which results in an average pH of 7.0 and an average PaCO₂ of 100 mmHg (Nahas, 1965). A
4
5 study by Longmore et al. showed increased fat synthesis in a perfused liver when the level of
6
7 CO₂ was raised in the medium (Longmore, 1967). This suggests that another factor adds to
8
9 the large increase in fat content found in the liver of guinea pigs exposed to 15% CO₂.
10
11 Similar changes have been observed in both guinea pigs and rats exposed to low levels of
12
13 CO₂ (3%).
14
15
16
17
18
19

20 **3. Pulmonary toxicity of carbon dioxide**

21 **a- Respiratory function**

22
23
24
25
26 Most toxicological studies have been focused on respiratory damages. Under normal
27
28 conditions, spontaneous breathing requires feedback controls in which detection of blood gas
29
30 levels and pH are critical. CO₂ sensing depends on central chemoreceptors (CCRs) located at
31
32 multiple sites. They are highly sensitive to CO₂, as an evident change in ventilation occurs
33
34 with an increase in PaCO₂ as small as 0.015% (1 mm Hg). Such sensitivity is likely to be
35
36 attributable to the inherent properties of CO₂/pH sensing molecules (mainly receptors and
37
38 channels) and their modulation in brainstem neuronal networks. Each of these molecules
39
40 covers a small range in the whole sensory spectrum. With multiple sensors arranged in
41
42 parallel, both high sensitivity and broad bandwidth may be achieved (Jiang, 2005).
43
44
45
46

47
48 CO₂ is an asphyxiant and loss of consciousness can occur when exposed to 30%
49
50 during one minute, or 10% during 5 to 10 minutes (NAP, 2007). The effects of hypercapnia
51
52 on the respiratory function appear immediately and at relatively low concentrations (from
53
54 1% CO₂). Following exposure to 5% carbon dioxide, there is an increased respiratory minute
55
56 volume, increased respiratory amplitude and frequency, as well as a decrease in the airway
57
58 conductance. In monkeys, the respiratory rate increased two-fold until a 10 % carbon dioxide
59
60 concentration was reached and thereafter decreased until animals died (Stinson, 1970). In a

1
2
3 human study (Schaefer, 1963), 23 healthy men were exposed at a constant level of 1.5 %
4 CO₂ in air for 42 days in a submarine which served as the experimental chamber.
5
6 Throughout the exposure to CO₂ the respiratory minute volume and alveolar CO₂ tension
7
8 were increased. During the post-exposure period (9 days), the respiratory minute volume
9
10 decreased event though the CO₂ tension remained elevated. The authors divided the 42-day
11
12 exposure period into two parts. The first phase (days 1-23, uncompensated acidosis) was
13
14 characterized by a significant increased of the alveolar carbon dioxide tension, carbon
15
16 dioxide excretion and respiratory exchange ratio. The second phase (days 24-42,
17
18 compensated acidosis) was characterized by an increased excretion of carbon dioxide.
19
20
21
22
23

24
25 The early acute response to high CO₂ tensions (above 5%) is characterized by an
26
27 enhanced respiratory volume. This is illustrated by the very rapid (within minutes) of Penh
28
29 value calculated from plethysmography-measured parameters during the exposure of mice to
30
31 5, 10 or 15% (Abolhassani 2009).
32
33

34 **b- Acute and chronic lung toxicity**

35
36
37 Douglas and al. studied the consequences of chronic exposure (up to 6 weeks) to 1%
38
39 CO₂ in guinea pigs (Douglas, 1979). They observed an elevation of PaCO₂ associated with a
40
41 metabolic acidosis that reach a maximum at four weeks of exposure and persist even after
42
43 two weeks of recovery in normal air. Electron microscopy analysis showed changes in cell
44
45 fine structure of type II alveolar pneumocytes (granular pneumocytes) and lamellar bodies,
46
47 hyperplasia (cluster of 2-4 cells) and hypertrophia of these cells as compared to control,
48
49 suggesting an increased activity of theses cells.
50
51
52
53

54
55 Animal studies have indicated that chronic exposure to higher level of CO₂ can cause
56
57 hyaline membrane formation and atelectasis in guinea pigs and can cause edema in rat lungs.
58
59 Niemoller and Schaefer (Niemoeller, 1962) exposed guinea pigs and rats to different CO₂
60
concentrations (from 3 to 15%) during prolonged and continuous exposures (from two days

1
2
3 to six months). Loss of surfactant (complex system of lipids, proteins and lipoproteins which
4 allows the alveoli to remain open throughout the normal cycle of inhalation and exhalation)
5 was associated with hyaline membrane (fibrins, cellular debris lining or filling the alveolar
6 spaces) formation that led to decreased gas exchange, associated with respiratory distress
7 syndrome. Microscopic examination indicated that guinea pigs exposed to 3 and 15% CO₂
8 developed hyaline membranes (respectively from fourth and first days), while those exposed
9 to 1.5% CO₂ did not, supporting the hypothesis of a threshold for CO₂ induced lung toxicity.
10
11
12
13
14
15
16
17
18
19

20 In a follow-up study (Schaefer, 1964a), guinea pigs were exposed to CO₂ and data
21 were gathered from electron microscope studies, surface tension measurements of lung
22 tissue, and additional histochemical studies. These authors identified four phases of
23 pulmonary changes caused by 15% carbon dioxide. The initial phase (6 hours) was marked
24 by uncompensated respiratory acidosis accompanied by pulmonary effusion (edema,
25 congestion, atelectasis and hemorrhage) as well as changes in the lamellar bodies
26 (intracellular stores of surfactant) of the granular (type II) pneumocytes. This period was not
27 associated with hyaline membrane formation. The second phase (6-24 hours) was associated
28 with hyaline membrane formation. During the third phase (days 2-7), the surface tension
29 returned to normal, the pulmonary edema diminished and hyaline membranes disappeared.
30 The final phase was one of recovery despite the fact that the pCO₂ remained elevated.
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45

46 Recently, Abolhassani demonstrated that inhalation of levels of carbon dioxide above
47 5 %, for one hour, induced pulmonary inflammation. The authors showed an increase in the
48 secretion of the pro-inflammatory cytokines TNF alpha, interleukin 8, interleukin 6 Mip1-
49 alpha as well as mucin 5AC, a major pulmonary mucus glycoprotein overexpressed during
50 inflammation. This inflammation was caused by the methylation of the C subunit of the
51 phosphatase PP2A, which in turn controls the translocation of the transcription factor NF-
52 κB. Interestingly, complementary *in vitro* experiments do not seem to correlate pH variations
53
54
55
56
57
58
59
60

1
2
3 to this inflammation response as IL-8 secretion was not induced in response to acidic pH
4 imposed in the culture medium. These molecular biological findings were confirmed by
5
6 microscopic examination of the lung. Extensive lung inflammation with infiltration of the
7
8 parenchyma by lymphocytes and monocytes was observed (Abolhassani, 2009).
9
10

11
12
13 Additionally, Ryu and al. compared the consequences of the exposure of mouse
14
15 neonates versus adults to 8% CO₂ for two weeks (Ryu, 2010). They showed that CO₂
16
17 exposure decreased lung alveolar walls thickness, reduces lung weight and alters lung matrix
18
19 proteic composition (among them decrease of interstitial collagen) in young mice. In
20
21 comparison, adult lungs were not affected, which highlighted the sensitivity of young
22
23 individuals to CO₂ tension variations.
24
25
26
27
28

29 30 **4. Effects of carbon dioxide on cardiovascular function**

31
32

33 The first noticeable effects of CO₂ inhalation is an increase in heart rate. For example,
34
35 exposure to levels of at least 5% CO₂ resulted in the first signs of cardiovascular and
36
37 vasomotor impacts (cardiac frequency and arterial pressure, peripheric vasodilatation) in
38
39 humans. The same signs are observed in dogs and monkeys (Stinson, 1970) at concentrations
40
41 of up to 10 % of the gas. In dogs with left ventricular failure (embolization of the left
42
43 coronary artery), hypercapnia aggravated the heart failure (increase of the left ventricular
44
45 end-diastolic pressure, mean right arterial pressure and mean right arterial pressure);
46
47 however, the pump function of the heart was unchanged (Wexels, 1987). A reversion of the
48
49 of the central hemodynamics changes was observed when pH is normalized during
50
51 hypercapnia, meaning that pH, and not PaCO₂, was responsible for the observed
52
53 hemodynamic deterioration.
54
55
56
57
58

59 Schaefer analyzed the heart histopathology of guinea pigs after exposure to 1.5, 3 or 15%
60
CO₂ (Schaefer, 1971). No evidence of permanent myocardial damage was seen either in

1
2
3 animals that expired during the period of acute acidosis or that were sacrificed at one day or
4
5 seven days following initiation of the exposure. However, a small amount of lipid (red O
6
7 stain) was seen in one animal after one day, and by seven days positive material was seen in
8
9 five of ten animals. To our knowledge, no similar experimentation was performed to
10
11 reproduce these data.
12
13
14

15 16 17 18 **5. Effects of carbon dioxide on central nervous system and neuroendocrine** 19 **function** 20

21
22 Carbon dioxide is a key factor in the control of respiration and cerebral circulation. It acts
23
24 peripherally, both as a vasodilator and as a vasoconstrictor, and is a powerful cerebral
25
26 vasodilator.
27
28

29
30 The majority of the studies reported that chronic low concentration of CO₂ induces low
31
32 to mild effects: visual impairment occurred at 1% CO₂ and headaches were noticed in the
33
34 first days of exposure above 2% (National Research Council [NRC], 2007). In general, no
35
36 specific neurobehavioral changes or adverse effects were reported with level at up to 4% for
37
38 duration for up to two weeks (Storm, 1974). However, more recent studies showed a
39
40 decrease in stereoacuity and a decrease in the ability to detect motion with levels above 2.5%
41
42 (Sun, 1996; Yang, 1997).
43
44
45

46
47 At high concentrations, CO₂ exerts a stimulating effect on the central nervous system,
48
49 while excessive levels exert depressant effects (Lambertsen, 1971). Exposure to 10 % carbon
50
51 dioxide during approximately 1.5 minutes causes neurologic signs including eye flickering,
52
53 psychomotor excitation, and myoclonic twitches. At 15%, the same signs were recorded, as
54
55 well as increased muscle tone, perspiration, flushing, restlessness, dilated pupils, leg flexion
56
57 and torsion spasms. Apart from excitability, no abnormal behavior has ever been observed
58
59 after carbon dioxide exposure (psychomotor tests, resolution of problems...) (NIOSH, 1976).
60

1
2
3
4 Carbon dioxide induces changes in the secretion of hormones. Continuous exposition
5
6 (15% CO₂, 7 days) (Schaefer, 1968) stimulates the adrenal gland of guinea pigs. If animals
7
8 are intermittently exposed to this same concentration of CO₂ (8 hours daily for 7 days), there
9
10 is an initial fall of pH but no compensation of respiratory acidosis and no changes of the
11
12 sympathoadrenal responses. From these observations, the authors suggest that the stress
13
14 response in chronic hypercapnia depends on extracellular and related intracellular changes
15
16 and is representative of a non-specific pH-dependent effect.
17
18
19
20
21

22 **6. Alteration of the reproductive capacity by carbon dioxide**

23
24
25

26 In rats (VanDemark, 1972), carbon dioxide causes degenerative changes of the testes.
27
28 These modifications depend on both the dose (2.5 %, 5% or 10% carbon dioxide) and the
29
30 duration of exposure (1 to 8 hours). Major histological effects included tubular disturbances
31
32 such as sloughing as well as loss of luminal definition (5% during 4 hours), degenerative
33
34 changes such as streaking and vacuolization (10% during 4 hours). These modifications are
35
36 reversible, as testes were normal 36 hours after carbon dioxide exposure.
37
38
39

40 A concentration of 15% chronic CO₂ affects the spermatogenesis of guinea pigs and rats
41
42 (Schaefer, 1971). The first changes in spermatogenesis are noted after 48 hours. There is a
43
44 marked decrease in the number of mature spermatozooids. After 3-7 days, multinucleated
45
46 giant cells are seen. On the other hand, prolonged exposure to low levels (1.5 and 3% CO₂)
47
48 did not produce any spermatogenic arrest in guinea pigs and rats. Surprisingly, there are no
49
50 recent data relating carbon dioxide exposure to fertility.
51
52
53
54
55

56 **7. Teratogenicity of carbon dioxide**

57
58
59
60

1
2
3
4 Hypercapnia is teratogenic. Exposition of rats to 6% carbon dioxide (single 24 -period
5
6 between days 5 and 21 of pregnancy) causes some malformations in the newborn pups
7
8 (Haring, 1960): cardiac malformations in 24% of the tested animals (7% in control), and
9
10 skeletal malformations in 11% (0.6% in control). Exposure to higher CO₂ levels (10 %) and
11
12 consecutive acidosis of neonatal rats promote the retinopathy of prematurity, a potentially
13
14 blinding eye disorder that primarily affects premature infants (Holmes, 1994 and 1998).
15
16

17
18 In rabbits (Grote, 1965) exposed to 10-13% carbon dioxide, the newborn pups had
19
20 vertebral malformations. Furthermore, Nagai A. et al. (Nagai, 1987) examined fetuses from
21
22 rabbits exposed from day 21 to day 28 of gestation to 8 % CO₂ for 8 hours each day. These
23
24 fetuses weighed less and presented numerous characteristics of increased tissue and cellular
25
26 maturation of the lung (increased distended lung volumes, increased volume proportion of
27
28 air spaces, decreased air-space wall, less glycogen and lamellar bodies ...).
29
30

31
32 Mice exposed to 20% CO₂ for 8 hours on day 10 of gestation produced right-sided
33
34 postaxial forelimb ectrodactyly in 23% of the offspring. Rather than metabolic acidosis, it
35
36 would seem that the primary teratogenic factor in hypercapnia is elevated CO₂ tension (low
37
38 incidence of ectrodactyly associated with NH₄Cl-induced acidosis). Moreover, there is a
39
40 strong correlation between maternal serum CO₂ content and the incidence of ectrodactyly
41
42 (Weaver, 1984).
43
44
45
46
47
48

49 **8. Carcinogenic potential of carbon dioxide**

50 **a- In vitro alteration of cell fate by CO₂ exposure**

51
52
53 In 1925, Bauer (Bauer, 1925) exposed chicken tissues to carbon dioxide
54
55 (concentration level not reported) *in vitro* during 6 to 8 days. The author primarily described
56
57 the consequences of carbon dioxide on dividing cells, and, in particular the retardation of cell
58
59 division (in prophase, anaphase and telophase) as well as some alterations of the division
60

1
2
3 process: “It was noted that the entire equatorial plate moved slowly from one pole to another
4
5 without a division of the chromosomes”.
6
7

8 In 1927, Mottram (Mottram, 1927) described that the CO₂ tension and/or acidity
9 applied to culture cells control cell activity, in particular cell migration. He also evoked the
10 role that carbon dioxide could play in cancer aetiology. In a follow-up experiment, Mottram
11 cultivated kidneys cells and fibroblasts of young rats with different concentrations of carbon
12 dioxide during 3 days; cells were then fixed and mitosis were counted (Mottram, 1928).
13 From these observations, he deduced that the optimum tension of CO₂ for the cell division in
14 normal cells is the physiological CO₂ tension (6% CO₂) but that cell division occurs at
15 concentrations above (up to 30% CO₂) and below (0 mmHg) this normal tension.
16 Interestingly, whilst counting these mitoses in fibroblasts, many abnormal features were
17 observed in the cultures grown at elevated CO₂ tensions. These features consisted of “an
18 irregular migration of the chromatin towards the centrosomes; some chromatin remained
19 suspended at the equator of the spindle, while other fragments had already migrated to the
20 centrosome. This unusual arrangement was more often than not asymmetrical, a fragment of
21 chromatin being present at one centrosome with none at the other, or more fragments at one
22 centrosome than at the other.[...] It was also observed that the size of the nuclei of
23 undividing cells under high tension of CO₂ was increased, while reduced at low tensions, as
24 compared to nuclei of cells at 40 mmHg (6%) CO₂.” Thus, high carbon dioxide
25 concentration clearly acts as a disrupter of normal cell division processes. The author also
26 noticed that these abnormalities were similar to those observed in cells that had been
27 subjected to X-irradiation, where, “besides fragmentation of the chromatin into fine granules,
28 delay in its migration to the centrosomes occurs, so that whilst some chromatin has moved to
29 the centrosomes, some remains suspended at the equator of the spindle”. Similarly as after
30 X-irradiation, an increase in size of the cell nuclei was observed under high CO₂
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 concentrations. These observations support the hypothesis of a role of supraphysiological
4 concentrations of carbon dioxide in carcinogenesis via the disruption of cell division
5 processes. Recent data partially complete these earlier findings.
6
7
8
9

10 Schuller et al. addressed the mechanisms of cell proliferation in response to nicotine and
11 NNK (nitrosamine 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone) in normal pulmonary
12 neuroendocrine cells (PNE) cells derived from fetal hamster lung, and in two cell lines
13 derived from human neuroendocrine lung cancers (Schuller, 1994). Their data demonstrated
14 that the mitogenic effects of nicotine and NNK are potentiated by elevated levels of CO₂
15 (from 8 to 12%) in a concentration dependant manner. Similarly, Merryman demonstrated
16 that a concentration of 10 % CO₂ stimulated the proliferation of small cell lung cancer cells
17 exposed *in vitro*. CO₂ activated the MAP kinase pathway and could be considered as an
18 important messenger molecule in the lung (Merryman, 1997). Interestingly, this article also
19 underlined that chronic nonneoplastic pulmonary diseases (COPD, asthma, emphysema,
20 chronic bronchitis) are characterised by an impaired respiration and an augmentation of
21 carbon dioxide pulmonary tension (7 to 40%) which might promote lung cancer
22 development.
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40

41 These articles supports the hypothesis that high carbon dioxide tension might promote
42 cancer development.
43
44
45
46
47
48
49

50 ***b-* In vivo carcinogenicity of carbon dioxide**

51
52 The following articles describe the carcinogenic effects of CO₂ *in vivo*. It should be
53 noted, however, that for most of them, the concentrations of carbon dioxide used were very
54 high.
55
56
57
58

59 In a transplantation experiment, skin autografts were exposed *in vitro* before
60 transplantation during 48 hours to 45 to 48% CO₂ in air (control: room air culture). Although

1
2
3 some malignant lymphoma were observed in host animals using untreated autografts, the
4
5 lymphoma incidence was highest in the recipients of the CO₂-treated grafts. Other
6
7 abnormalities of the reticuloendothelial system were noted: proliferation of lymph follicles
8
9 into irregular masses of pleomorphic cells, hyperplasia with concomitant atrophy of the
10
11 lymphoid tissue, and replacement of the lymph follicles by malignant lymphoid cells
12
13 (Goldsmith, 1975).
14
15

16
17
18 The long-term clinical effects of high CO₂ tensions on various normal tissues in mice
19
20 have been investigated (Goldsmith, 1980). Different tissues were exposed *in vitro* to a high
21
22 CO₂ proportion in air (45% CO₂) before transplantation into syngeneic or autologous hosts
23
24 or, in a second protocol, intraperitoneal tissues were exposed *in vivo* to CO₂-infusion
25
26 (99.99%), thus avoiding graft-host interactions. In the autologous grafts, pretreatment by
27
28 intraperitoneal CO₂-infusion induced lymphoma (60% incidence), air-infusion did not. Non-
29
30 lymphoid grafts exposed *in vitro* to elevated CO₂ induced only lymphoid malignancies. But
31
32 non-lymphoid tissues exposed *in vivo* to elevated CO₂ developed tumors of other tissues,
33
34 such as lung tumor, in addition to lymphoid malignancies. In fact, the spontaneous
35
36 pulmonary adenocarcinoma incidence doubles in the mice exposed to intraperitoneal CO₂.
37
38 The same morphological lymphoid abnormalities occurred in all lymphoma-developing
39
40 animals in these three experimental models: hyperplasia in the splenic T-cell areas appeared
41
42 most frequently (70-75 % incidence), whereas atrophy in T-cell areas of the lymph nodes and
43
44 B-cell areas hyperactivity were far less frequent.
45
46
47
48
49

50
51 In a mouse model for multiple laparoscopies, intraperitoneal insufflations of
52
53 approximately 3.5 ml of CO₂ were given daily to three groups of BALB/c mice for 11, 20,
54
55 and 32 consecutive days (control: air insufflation). Proliferation of splenic T-lymphocytes
56
57 (doubling of the T-cells spleen percentage) was an early, but transitory, immunologic
58
59 reaction in the spleen to intraperitoneal CO₂ insufflation. This was correlated to the late
60

1
2
3 occurrence of a high incidence of malignant lymphoma (approximately 60%). The long-term
4 survivors of CO₂ insufflation also developed a wide spectrum of malignancies that were not
5
6 of lymphoid origin, specifically adenocarcinoma in various organs: lung, kidney, adrenals,
7
8
9
10 ovary, gastrointestinal tract and salivary gland (Goldsmith, 1981).
11

12
13 The effects of high concentrations of CO₂ on experimental murine neuroblastoma
14
15 tumors have also been studied (West, 1978). The local growth of this neuroblastoma model
16
17 was not affected by concentrations of 76% and 55% of CO₂ applied for 10 and 30 minutes.
18
19 Although, the tumor bearing animals exposed to different CO₂ concentrations tended to
20
21 develop metastases more frequently than the control groups.
22
23
24
25
26

27 **9. Does carbon dioxide contribute to cigarette smoke toxicity?**

28
29

30
31 Below its immediately lethal concentration, carbon dioxide has long been considered as a
32
33 neutral compound for the body. However, recent studies raised interest on carbon dioxide in
34
35 relationship with chronic and/or intermittent long term exposure conditions that might
36
37 induce pathologic states, in particular nasal inflammation (Buron, 2009; Hacquermand,
38
39 2010) and pulmonary inflammation (Krohn, 2003; Abolhassani, 2009).
40
41

42
43 There are various situations when pCO₂ can rise in the inhaled air. First, during
44
45 professional exposures such as recurrent manipulation of dry ice, food and floral
46
47 preservation, wearing of mask, spacecraft, aircraft, submarine, mask, altitude, exposure to
48
49 combustion gas (Roberge, 2010; NRC, 2007; NRC, 2008). Second during pathological
50
51 exposures such as sleep apnea, pulmonary diseases (e.g. COPD, asthma, emphysema,
52
53 chronic bronchitis) (Windisch, 2005).
54
55

56
57 Smoking, as every combustion produces CO₂ (about 13% in the mainstream smoke).
58
59 And, even if the exposure is quite short, it is also repetitive and regular. Our hypothesis is
60
that carbon dioxide cigarette smoke content might be implicated in smokers pathologies.

1
2
3
4 Like CO₂, exposure to tobacco causes both acute and chronic lung inflammation
5 (BPCO). In a recent paper, we demonstrated that the acute toxicity of cigarette smoke is due
6 to carbon dioxide inhalation. Using a KOH filter, we were able to selectively trap
7 bicarbonates (and thus CO₂) but not oxygen or particles. The decrease in carbon dioxide
8 concentration results in the almost complete disappearance of the inflammatory syndrome
9 caused by cigarette smoking and expressed by the mice (Schwartz, 2010). The
10 precancerisation events described with *in vitro* exposure (cf. § 8) and the concept of cancer
11 as an inflammation-based disease lead us to think that carbon dioxide might be a major
12 contributor of lung carcinogenesis.
13
14

15
16 Bladder cancer is one of the most common human cancers, consisting about 6% to 2% of
17 all cancers among men and women, respectively. The exact mechanism by which smoking
18 increases the incidence of this malignancy is not known but some authors discuss the role
19 that inflammation might play (Burin, 1995). As hypercapnia, smoking tends to increase urine
20 acidity (Fix, 1986; Dales, 1978). Wald et al measured urinary pH in 145 cigarettes smokers
21 and found that 70% of the smokers have a pH value below 7 (Wald, 1984). Furthermore,
22 there is evidence that acidic urine has a negative influence on urothelial cell DNA adducts
23 levels in humans who have been exposed to benzidine, and the aromatic amines derived
24 from cigarette smoke. Individuals with urine pH lower than 6 had 10-fold higher DNA
25 adduct levels compared to subjects with urine pH 7 or greater (Rothman, 1997). Hence, the
26 acidity of urine caused by smoking might be an important susceptibility factor for the
27 development of bladder cancer.
28
29

30
31 Another aspect of carbon dioxide potential role in carcinogenesis is its implication in cell
32 metabolism. As previously described in §2, CO₂ is a key regulator of three metabolic
33 reactions. It can be presume that enhancing CO₂ levels in the cells will displace the
34 equilibrium of these reactions and favor the production of building elements for new cells.
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Smoking is known to be carcinogenic. Smoking induces a wide array of cancers such as
4 cancer of the lung, head and neck, esophagus and the bladder. The toxicity of carbon dioxide
5 has been established for close to a century. It is possible that its role in the epidemic of
6 tobacco related illnesses has been overlooked. Carbon dioxide could also interact with tar
7 making the animal more susceptible to its toxicity. There are no clear data demonstrating the
8 carcinogenicity of carbon dioxide, but short term exposures of cells or rodents suggest that it
9 is highly likely, a strong probability that further studies would confirm.
10
11
12
13
14
15
16
17
18
19
20
21

22 Acknowledgments

23 We acknowledge the help of Jean-Marc Steyaert and Edward Sanders. This work was funded
24 by Biorébus.
25
26
27
28
29
30
31

32 Competing interests

33 AG is an employee of Biorébus. The other authors declare that they have no competing
34 interests.
35
36
37
38
39
40
41
42
43
44
45

46 Bibliography

47
48
49
50
51
52 Abolhassani M, Guais A, Chaumet-Riffaud P.,Sasco A, Schwartz L (2009). Carbon dioxide
53 inhalation causes pulmonary inflammation. *Am J Physiol Lung Cell Mol Physiol* 296:L657-
54 665.
55
56

57
58 Alora-Pali MB, Perkins AC, Van Cott A, Kimball AB (2010). Efficacy and tolerability of a
59 cosmetically acceptable coal tar solution in the treatment of moderate plaque psoriasis. *Am*
60 *J. Dermatol* 11:275-283.

- 1
2
3
4 Baggott, J. (1982). Gas transport and pH regulation. In Devlin TM, ed. Textbook of
5 Biochemistry with Clinical Correlations. New York: John Wiley and Sons, Pp. 1098-1123.
6
7
8 Bauer J. (1925). The effect of carbon dioxide on cells in tissue cultures. Bull Johns Hopkins
9 Hosp 37:420-439.
10
11
12 Bertoni G, Ciuchini C, Tappa R. (2004). Measurement of long-term average carbon dioxide
13 concentrations using passive diffusion sampling. Atmospheric Environment 38:1625-1630.
14
15
16 Briggs NC, Hall HI, Brann EA, Moriarty CJ, Levine RS.(2002). Cigarette smoking and risk
17 of Hodgkin's disease: a population-based case-control study. Am J Epidemiol 156:1011-
18 1020.
19
20
21
22 Brown EB Jr, Miller F. (1952). Ventricular fibrillation following a rapid fall in alveolar
23 carbon dioxide concentration. Am J Physiol 169:56-60.
24
25
26 Burin GJ, Gibb HJ, Hill RN.(1995). Human bladder cancer: evidence for a potential
27 irritation-induced mechanism. Food Chem Toxicol 33:785-795.
28
29
30 Buron G, Hacquemand R, Pourie G, Brand G. (2009). Carbon dioxide effects on olfactory
31 functioning: Behavioral, histological and immunohistochemical measurements. Toxicol Lett
32 188:251-257.
33
34
35
36 Burns D. (2003). Epidemiology of smoking-induced cardiovascular disease. Prog
37 Cardiovasc Dis 46:11-29.
38
39
40 Carter NW. (1959). Tissue and renal response to chronic respiratory acidosis. J Clin Invest
41 38: 949-960.
42
43
44 Cox PM, Betts RA, Jones CD, Spall SA, Totterdell IJ. (2000). Acceleration of global
45 warming due to carbon-cycle feedbacks in a coupled climate model. Nature 408:184-187.
46
47
48 Craig FN, Beecher HK. (1943). The effect of carbon dioxide tension on tissue metabolism
49 (retina). J Gen Physiology 26:473-478.
50
51
52 Dales LG, Friedman GD, Siegelau AB, Seltzer CC, Ury HK.(1978). Cigarette smoking
53 habits and urine characteristics: urinalysis abnormalities are more common in smokers, but
54 the reasons are unclear. Nephron 20:167-170.
55
56
57
58 Douglas WHJ, Schaefer KE, Messier AA, and Pasquale SM. (1979). Proliferation of
59 pneumocyte II cells in prolonged exposure to 1% CO₂. Undersea Biomed Res (Submarine
60 Suppl.):S135-S142.

1
2
3
4 Dube MF, Green CR. (1982). Methods of collection of smoke for analytical purposes. *Rec*
5 *Adv Tob Sci* 8:42–102.

6
7 Ezzati M, Henley SJ, Lopez AD, Thun MJ.(2005). Role of smoking in global and regional
8 cancer epidemiology: current patterns and data needs. *Int J Cancer*.116:963-971.

9
10
11 Fix AJ, Daughton D, Issenberg P.(1986). Cigarette smoking, urinary acidity, and nicotine
12 excretion under natural conditions. *Percept Mot Skills* 63:65-6.

13
14
15 Forster RE, Dogson SJ (2000). Membrane transport and provision of substrates for carbonic
16 anhydrase in vertebrates. In Chegwidde WR, Carter ND, Edwards YM Eds. *The carbonic*
17 *anhydrases: new horizons*. Basel: Birckhäuser, 263-280.

18
19
20
21 Glatte HA. (1967a). Carbon dioxide tolerance: a review. Review 5-67 USAF School of
22 Aerospace Medicine, Aerospace Medical Division, Brooks Air Force Base, Texas, USA.

23
24
25 Glatte HA. (1967b). Carbon dioxide tolerance studies. USAF School of Aerospace
26 Medicine, Aerospace Medical Division, Brooks Air Force Base, Texas, USA.

27
28
29 Goldsmith AE, Narvaez R. (1975). Lymphomas as sequelae of the transplantation of CO₂-
30 treated skin autografts in mice. *Oncology* 32:247-265.

31
32
33 Goldsmith AE, Ryan GF, Joseph AB. (1980). Metabolic carcinogenesis-induction of murine
34 lymphoma by CO₂-treatment *in vivo* and *in vitro*. *Jpn J Med Sci Biol* 33:7-18.

35
36
37 Goldsmith AE, Ryan GF, Joseph AB. (1981). A murine model for multiple laparoscopies: I.
38 Early lymphoid response to intraperitoneal insufflation of CO₂. *Cancer Detect Prev* 4:109-
39 115.

40
41
42
43 Grote W. (1965). Disturbances of embryonic development at elevated CO₂ and O₂ partial
44 pressure at reduced atmospheric pressure. *Z Morphol Anthropol* 56:165-94.

45
46
47 Hacquemand R, Buron G, Pourie G, Karrer M, Jacquot L, Brand G. (2010). Effects of CO₂
48 inhalation exposure on mice vomeronasal epithelium. *Cell Biol Toxicol* 26:309-317.

49
50
51 Haring OM. (1960). Cardiac malformation in rats induced by exposure of the mother to
52 carbon dioxide during pregnancy. *Circ Res* 8:1218-1227.

53
54
55 Harris JE, Thun MJ, Mondul AM, Callee EE. (2004). Cigarette tar yields in relation to
56 mortality from lung cancer in CPS II. *BMJ* 328:7431-7432.

1
2
3
4 Hollidge-Horvat MG, Parolin ML, Wong D, Jones NL, Heigenhauser GJ.(1999). Effect of
5 induced metabolic acidosis on human skeletal muscle metabolism during exercise. *Am J*
6 *Physiol* 277:E647-58.

7
8
9 Holmes JM, Duffner LA, Kappil JC.(1994). The effect of raised inspired carbon dioxide on
10 developing rat retinal vasculature exposed to elevated oxygen. *Curr Eye Res* 13:779-782.

11
12 Holmes JM, Zhang S, Leske DA, Lanier WL. (1998). Carbon dioxide-induced retinopathy in
13 the neonatal rat. *Curr Eye Res* 17:608-616.

14
15
16 Keeling CD, Whorf TP, Walhen M, van der Plicht. (1995). Interannual extremes in the rate
17 of rise of atmospheric carbon dioxide since 1980. *Nature* 375:666-670.

18
19
20 Krohn TC, Kornerup Hansen A, Dragsted N. (2003). The impact of low levels of carbon
21 dioxide on rats. *Laboratory Animals* 37:94-99.

22
23
24 Lahiri S, Forster RE 2nd. (2003). CO₂/H(+) sensing: peripheral and central chemoreception.
25 *Int J Biochem Cell Biol.* 35:1413-1435.

26
27
28
29 Lambertsen CJ. (1971). Therapeutic gases- oxygen, carbon dioxide and helium, in Di Palma
30 JR (ed). *Drill's Pharmacology in Medicine*, ed 4. New York: McGraw-Hill Book Co, chap
31 55.

32
33
34
35 Lang CJ, Dong P, Hosszu EK, Doyle IR. (2005). Effect of CO₂ on LPS-induced cytokine
36 responses in rat alveolar macrophages. *Am J Physiol Lung Cell Mol Physiol* 289:L96-L103.

37
38
39
40 Leboeuf RA, Kerchaert GA. (1987). Enhanced morphological transformation of early
41 passage Syrian hamster embryo cells cultured in medium with a reduced bicarbonate
42 concentration and pH. *Carcinogenesis* 8:689-697.

43
44
45 Longmore WJ, Hastings AB, Harrison E, Liem HH. (1967). Effect of CO₂ and cations on
46 fatty acid and cholesterol synthesis by liver in vitro. *Am J Physiol* 212:221-227.

47
48
49 Man LX, Chang B.(2006). Maternal cigarette smoking during pregnancy increases the risk
50 of having a child with a congenital digital anomaly. *Plast Reconstr Surg* 117:301-308.

51
52
53 Mattson SN, Calarco KE, Chambers CD, Jones KL.(2002). Interaction of maternal smoking
54 and other in-pregnancy exposures: analytic considerations. *Neurotoxicol Teratol* 24:359-367.

55
56
57
58 Mekhail K, Gunaratnam L, Bonicalzi ME, Lee S. (2004). HIF activation by pH-dependent
59 nucleolar sequestration of VHL. *Nat Cell Biol.* 6:642-647.

- 1
2
3
4 Merryman JI, Park PG, Schuller HM. (1997). Carbon dioxide, an important messenger
5 molecule for small cell lung cancer. *Chest* 112:779-784.
6
7
8 Mottram J.C. (1927). The role of carbon dioxide in the growth of normal and tumour cells.
9
10 The *Lancet*. 210:1232-1234.
11
12 Mottram JC. (1928). On the division of cells under varying tensions of carbon dioxide. *Br J*
13 *Exp Path* 9:240-244.
14
15 Mukherjee DP, Sing SP. (1967). Effect of increased carbon dioxide in inspired air on the
16 morphology of spermatozoa and fertility of mice. *J Reprod Fertil* 13:165-167.
17
18 Nahas G.G., Poyart C.F. (1965). Effect of arterial pH alteration on metabolic activity of
19 norepinephrine. *Am J Physiol*. 213:765-772.
20
21
22 Nagai A, Thurlbeck WM, Deboeck C, Ioffe S, Chernick V. (1987). The effect of maternal
23 CO₂ breathing on lung development of fetuses in the rabbit. Morphologic and morphometric
24 studies. *Am Rev Respir Dis* 135:130-6.
25
26
27 Neese RA, Benowitz NL, Hoh R, Faix D, LaBua A, Pun K, Hellerstein MK. (1994).
28 Metabolic interactions between surplus dietary energy intake and cigarette smoking or its
29 cessation. *Am J Physiol*. 267:E1023-34.
30
31
32 Nelson E, Jodscheit K, Guo Y. (1999). Maternal passive smoking during pregnancy and fetal
33 developmental toxicity. Part 1: gross morphological effects. *Hum Exp Toxicol* 18:252-6.
34
35
36 Niemoeller H, Schaeffer KE. (1962). Development of hyaline membrane and atelectases in
37 experimental chronic respiratory acidosis. *Proc Soc Exp Biol Med* 110:804-808.
38
39
40 NIOSH (1976). Criteria for a recommended standard. Occupational exposure to carbon
41 dioxide. Washington US department of Health, Education and Welfare, PuWc Health
42 Service.
43
44
45 NRC. (2007). Carbon dioxide. In *Emergency and Continuous Exposure Guidance Levels for*
46 *Selected Submarine Contaminants, Vol.1*. Washington, DC: The National Academies Press,
47 46-66.
48
49
50 NRC. (2008). Carbon dioxide. In *Spacecraft Maximum Allowable Concentrations for*
51 *Selected Airborn Contamnants, Vol.5*. Washington, DC: The National Academies Press,
52 112-124.
53
54
55
56
57
58
59
60

- 1
2
3
4 Ono Y, Lin L, Storey BT, Taguchi Y, Dodgson SJ, Forster RE. (1996). Continuous
5 measurement of $^{13}\text{C}16\text{O}_2$ production from ^{13}C pyruvate by intact liver mitochondria:
6 effect of HCO_3^- . *Am J Physiol* 270:C98-106.
7
8
9 Poyart CF, Nahas GG, Vulliemoz Y. (1968). Inhibition of activated lipolysis by acidosis.
10 *Mol Pharmacol* 4:389-401.
11
12
13 Pringle PJ, Geary MP, Rodeck CH, Kingdom JC, Kayamba-Kay's S, Hindmarsh PC. (2005).
14 The influence of cigarette smoking on antenatal growth, birth size, and the insulin-like
15 growth factor axis. *J Clin Endocrinol Metab* 90:2556-2562.
16
17
18 Roberge RJ, Coca A, Williams WJ, Powell JB, Palmiero AJ. (2010). Physiological impact of
19 the N95 filtering facepiece respirator on healthcare workers. *Respir Care* 55:569-577.
20
21
22
23 Rothman N, Talaska G, Hayes RB, Bhatnagar VK, Bell DA, Lakshmi VM, Kashyap
24 SK, Dosemeci M, Kashyap R, Hsu FF, Jaeger M, Hirvonen A, Parikh DJ, Davis BB, Zenser
25 TV. (1997). Acidic urine pH is associated with elevated levels of free urinary benzidine and
26 N-acetylbenzidine and urothelial cell DNA adducts in exposed workers. *Cancer Epidemiol*
27 *Biomarkers Prev* Dec 6:1039-1042.
28
29
30
31
32
33 Ryu J, Heldt GP, Nguyen M, Gavrialov O, Haddad GG (2010). Chronic hypercapnia alters
34 lung matrix composition in mouse pups. *J Appl Physiol* 109:203-210.
35
36
37
38 Schaefer KE, Hasson M, Niemoller H. (1961). Effect of prolonged exposure to 15% CO_2 on
39 calcium and phosphorus metabolism. *Proc Soc Exp Biol Med* 107:355-359.
40
41
42
43 Schaefer KE, Hastings BJ, Carey CR, Nichols G Jr. (1963). Respiratory acclimatization to
44 carbon dioxide. *J Appl Physiol* 18: 1071-1078.
45
46
47
48 Schaefer KE, Avery ME, Bensch K. (1964a). Time course of changes in surface tension and
49 morphology of alveolar epithelial cells in CO_2 -induced hyaline membrane disease. *J Clin*
50 *Invest* 43:2080-2093.
51
52
53
54 Schaefer KE, Nichols G Jr, Carey CR. (1964b). Acid-base balance and blood and urine
55 electrolytes of man during acclimatization to CO_2 . *J Appl Physiol* 19: 48-58.
56
57
58
59 Schaefer KE, McCabe N, Withers J. (1968). Stress response in chronic hypercapnia. *Am J*
60 *Physiol* 214:543-548.
- Schaefer KE. (1970). The effect of intermittent exposure to 3 % CO_2 on acid-base balance and electrolyte excretion, report N° 635. Groton, Conn, US Navy Dept, Bureau of Medicine

1
2
3 and Surgery, Naval Submarine Medical Center, Submarine Medical Research Laboratory, 1-
4
5 8.

6
7 Schaefer KE (1971). Chronic CO₂ toxicity: species difference in physiological and
8
9 histopathological effects. report N° 656. Groton, Conn, US Navy Dept, Bureau of Medicine
10
11 and Surgery, Naval Submarine Medical Center, Submarine Medical Research Laboratory, 1-
12
13 26.

14
15 Schuller HM. (1994). Carbon dioxide potentiates the mitogenic effects of nicotine and its
16
17 carcinogenic derivative, NNK, in normal and neoplastic neuroendocrine lung cells via
18
19 stimulation of autocrine and protein kinase C-dependent mitogenic pathways.
20
21 Neurotoxicology 15: 877-886.

22
23 Schwartz L, Guais A, Chaumet-Riffaud P, Grevillot G, Sascio A., Molina T, Abolhassani M.
24
25 (2010). Carbon dioxide is largely responsible for the acute inflammatory effects of tobacco
26
27 smoke. Inhal Toxicol 22: 543-551.

28
29 Shusterman DJ, Balmes JR. (1997). A comparison of two methods for determining nasal
30
31 irritant sensitivity. Am J Rhinol 11:371-378.

32
33 Smith RM (2005). Evaluation of Arterial Blood Gases and Acid-Base homeostasis. In
34
35 Bordow RA, Ries AL, Morris T A ed. Manual of Clinical Problems in Pulmonary medicine.
36
37 Philadelphia: Lippincott Williams & Wilkins, 28-36.

38
39 Stinson JM, Mattsson JL. (1970). Tolerance of rhesus monkeys to graded increase in
40
41 environmental CO₂- Serial changes in heart rate and cardia rhythm. Aerosp Med 42:78-80.

42
43 Storm, WF, CL Giannetta. (1974). Effects of hypercapnia and bedrest on psychomotor
44
45 performance. Aerosp Med 45:431-433.

46
47 Sun M, Sun C, Yang Y. (1996). Effect of low-concentration CO₂ on stereoacuity and energy
48
49 expenditure. Aviat Space Environ Med 67:34-39.

50
51 Talamini R, Polesel J, Montella M, Maso LD, Crispo A, Spina M, Franceschi S, Crovatto M,
52
53 La Vecchia C. (2005). Smoking and non-Hodgkin lymphoma: case-control study in Italy. Int
54
55 J Cancer 115:606-610.

56
57 Triner L, Nahas G. (1965). Acidosis: effect on lipolytic activity of norepinephrine in isolated
58
59 fat cells. Science 150:1725-7.
60

1
2
3 VanDemark NL , Schanbacher BD, Gomes WR. (1972). Alterations in testes of rats exposed
4 to elevated atmospheric carbon dioxide. J Reprod Fertil 28: 457-459.
5

6
7 Vitkus SJ, Hanifin SA, McGee DW. (1998). Factors affecting Caco-2 intestinal epithelial
8 cell interleukin-6 secretion. *In vitro* Cell Dev Biol Anim 34:660-664.
9

10
11 Wald NJ, Idle M, Boreham J, Bailey A, Van Vunakis H (1984). Urinary nicotine
12 concentrations in cigarette and pipe smokers. Thorax 39:365-368.
13

14
15 Warburg O. (1924). Ueber den Stoffwechsel der Tumoren; Biochemische Zeitschrift.
16 152:319-344.
17

18
19 Weaver TE, Scott WJ Jr. (1984). Acetazolamide teratogenesis: interaction of maternal
20 metabolic and respiratory acidosis in the induction of ectrodactyly in C57BL/6J mice.
21 Teratology 30:195-202.
22

23
24 West CR. (1978). A study of the effect of CO₂ on experimental neuroblastoma. Res
25 Commun Chem Pathol Pharmacol 20:399-404.
26

27
28 West MA, Baker J, Bellingham J. (1996). Kinetics of decreased LPS-stimulated cytokine
29 release by macrophages exposed to CO₂. J Surg Res 63:269-274.
30

31
32 Wexels JC, Mjøs OD. (1987). Effects of carbon dioxide and pH on myocardial function in
33 dogs with acute left ventricular failure. Crit Care Med 15:1116-1120.
34

35
36 Windisch W, Kostić S, Dreher M, Virchow JC Jr, Sorichter S. (2005). Outcome of patients
37 with stable COPD receiving controlled noninvasive positive pressure ventilation aimed at a
38 maximal reduction of Pa(CO₂). Chest 128:657-662.
39

40
41 Wood SC. (1978). Regulation of intracellular pH in lungs and other tissues during
42 hypercapnia. J Appl Physiol 45:115-118.
43

44
45 Yang Y, Sun C, and Sun M. (1997). The effect of moderately increased CO₂ concentration
46 on perception of coherent motion. Aviat Space Environ Med 68:187-191.
47
48
49
50
51
52
53
54
55
56
57
58
59
60